

Review of the European *Greenomyia* Brunetti (Diptera, Mycetophilidae) with new descriptions of females

Olavi Kurina¹, Kjell Hedmark², Mats Karström³, Jostein Kjærandsen⁴

1 Institute of Agricultural and Environmental Sciences, Estonian University of Life Sciences, Riia st 181, 51014 Tartu, Estonia **2** Kungsfågelvägen 1C, S- 79432 Orsa, Sweden **3** Älrvägen 4, S-96030 Vuollerim, Sweden **4** Department of Biology, Museum of Zoology, Lund University, Helgonavägen 3, S-223 62 Lund, Sweden

Corresponding author: Olavi Kurina (olavi.kurina@emu.ee)

Academic editor: Chris Thompson | Received 12 April 2010 | Accepted 13 January 2011 | Published 26 January 2011

Citation: Kurina O, Hedmark K, Karström M, Kjærandsen J (2011) Review of the European *Greenomyia* Brunetti (Diptera, Mycetophilidae) with new descriptions of females ZooKeys 77: 31–50. doi: 10.3897/zookeys.77.936

Abstract

The females of the four continental *Greenomyia* Brunetti species in Europe are associated with the males, diagnosed and keyed, providing the first association and description of the females of *G. baikalica* Zaitzev, 1994 and *G. stackelbergi* Zaitzev, 1982. Colour photographs of their habitus and line drawings of their female terminalia are provided. *Greenomyia mongolica* Laštovka & Matile, 1974 is found to be a senior synonym of *Greenomyia theresae* Matile, 2002. **syn. n.** The diagnostic characters used to distinguish between *Greenomyia* and *Neoclastobasis* Ostroverkhova in keys did not hold up to a closer scrutiny and leave the status of *Neoclastobasis* as separate genus questionable.

Keywords

Mycetophilidae, *Greenomyia*, *Neoclastobasis*, fungus gnats, systematics, identification key, synonymy, Europe

Introduction

The genus *Greenomyia* Brunetti was erected to distinguish a single oriental species, *Greenomyia nigricoxa* Brunetti, 1912. Since then several species (mainly Holarctic) have been described and new combinations proposed. Laštovka and Matile (1974) and Chandler and Ribeiro (1995) characterised the genus and Matile (2002) provided a key to all 11 world species of *Greenomyia*. Species of *Greenomyia* are mostly dark coloured medium-sized fungus gnats with a typical wing venation similar to that of *Leia*

Meigen, 1818 and allied genera where R_1 is notably shorter than the long and nearly horizontally aligned crossvein *rm*. Further diagnostic characters include the lateral ocelli being well separated from the eye margins and all veins reaching the wing margin (cf. Sølvi et al. 2000). Edwards (1925) introduced the tribe Leiini for *Leia*, *Greenomyia* and a number of other genera with an intermediate position between the subfamilies Sciophilinae and Mycetophilinae. Later the tribe has sometimes been given subfamily status (see review by Gammelmo 2004), but recent morphological and molecular studies have questioned its monophyly (e.g. Amorim and Rindal 2007; Rindal et al. 2009). *Greenomyia* appears most closely related to the genus *Neoclastobasis* Ostroverkhova, 1970, a genus that includes two eastern Palaearctic (*N. sibirica* Ostroverkhova, 1970 and *N. kamijoi* Sasakawa, 1964) and one European species (*N. draskovitsae* Matile, 1978). The genus *Neoclastobasis* is diagnosed by a prolonged apical palpal segment, the veins M_2 and CuA_1 terminating before the wing margin, and distinctive terminalia (Ostroverkhova 1970; Matile 1978; Zaitzev 1994; Sølvi et al. 2000).

Species of *Greenomyia* are not frequently encountered in Europe and apart from the widespread *G. mongolica* Laštovka & Matile, 1974 (e.g. Bechev 1989; Ševčík and Martinovský 1999; Plassmann 1996; Caspers 1996; Kurina 1997; Chandler 2010), they are generally thought of as quite rare. A revitalized focus on fungus gnats the last two decades, however, has yielded a number of new records and five species are now reported from Europe: viz. *G. baikalica* Zaitzev, 1994, *G. borealis* (Winnertz, 1863), *G. lucida* (Becker, 1908), *G. mongolica* (= *G. theresae* Matile, 2002, syn. n.), and *G. stackelbergi* Zaitzev, 1982 (see Fig. 29 for published records, except *G. lucida*, which is endemic to the Canary Islands).

While the males of most *Greenomyia* species are adequately illustrated and keyed (Zaitzev 1982, 1994; Laštovka and Matile 1974; Chandler and Ribeiro 1995; Matile 2002) the females of several species remain to be properly diagnosed and described. The current communication was initiated by the finding of three *Greenomyia* species from two localities 1 km apart in Vuollerim, Lule Lappmark in northern Sweden (Kjærandsen et al. 2007). The material gave us the opportunity to associate and describe females of two species for the first time. The shifted focus to females further revealed that the generic characters separating *Neoclastobasis* from *Greenomyia* do not hold, and highlights a need to re-evaluate their status as separate genera.

Material and methods

Material and collections from a wide range of Palaearctic sources were studied. The collecting methods, if known, are referred to in case by each specimen in the studied material section below. The following codens obtained from Evenhuis (2009) are used for depositories:

Coll. Hedmark Private collection of Kjell Hedmark, Orsa, Sweden.

Coll. Selin Private collection of Allan Selin, Tallinn, Estonia.

EIHU Hokkaido University Museum, Sapporo, Japan.

IZBE	Institute of Agricultural and Environmental Sciences, Estonian University of Life Sciences (former Institute of Zoology and Botany), Tartu, Estonia.
MNHN	Museum National d'Histoire Naturelle, Paris, France.
MZLU	Museum of Zoology, Lund University, Lund, Sweden.
ZIN	Zoological Institute of Russian Academy of Sciences, St. Petersburg, Russia.
ZMHB	Museum für Naturkunde Humbolt-Universität zu Berlin, Germany.
ZSM	Zoologische Staatssammlung in München, Germany

Three of four species were photographed and figured based on material collected in Vuollerim (Sweden), while illustrations of *G. mongolica* were based on Greek material. The terminalia were detached and cleared in a solution of KOH, followed by neutralization in acetic acid and washing in distilled water or in alcohol. The remaining chitinous parts were inserted into glycerine for detailed study, including illustration, and thereafter preserved as glycerine preparations in polyethylene micro vials. Habitus and wing photos were taken of specimens in alcohol, using a Canon 7D camera fitted with a Canon MP-E65 (F2.8 1–5 ×) lens. Illustrations of the terminalia were prepared using a U-DA drawing tube attached to a Olympus CX31 compound microscope. Terminalia are figured in three different positions: laterally, dorsally and ventrally. Sternite VIII was detached and figured separately to better expose the shape of hypoproct and gonapophysis IX. The preservation method of the studied specimens is indicated in the material section for each species. We used a 70–80 % solution of ethanol for alcohol preservation and the chemical method described by Vockeroth (1966) for dry-mounting from alcohol. Slide mounting in Euparal followed the method described by Kurina (2008b). Morphological terminology follows Söli (1997).

Systematics

Key to females of European *Greenomyia* species

Compiled from Zaitzev (1982, 1994), Chandler and Ribeiro (1995), Matile (2002) and original data.

- | | |
|---|---|
| 1 | Wing hyaline (Fig. 12). Coxae yellow, femora and tibiae yellow, only hind femur brown in apical fourth. Mesonotum yellow with three fused longitudinal brown stripes. Cercus one segmented. Sternite VIII with three apical incisions (Fig. 28) <i>G. stackelbergi</i> Zaitzev, 1982 |
| – | Wing with apical or preapical dark band. Mesonotum dark brown to black. Coxae yellow or blackish. Cercus one- or two-segmented. Sternite VIII with single central incision apically (Figs 25–27) 2 |
| 2 | Wing tip darkened on about apical third (Fig. 11) 3 |
| – | Wing with preapical dark band leaving tip hyaline (Figs 9–10) 4 |

- 3 Mid and hind coxae brown to black (Fig. 3). Cercus clearly two-segmented...
 ***G. mongolica* Laštovka & Matile, 1974**
- All coxae yellow or slightly darkened basally
 ***G. lucida* (Becker, 1908)** [Endemic to the Canary Islands, not seen]
- 4 Last palpal segment elongated (Fig. 6). C terminating distinctly before apex
 of wing, making R_5 straight to slightly sinuate (Fig. 10). Cercus two-segment-
 ed but segments partly fused (Figs 14, 18)
 ***Greenomyia borealis* (Winnertz, 1863)**
- Last palpal segment not elongated (Fig. 5). C terminating almost at apex of
 wing, making R_5 distinctly arched (Fig. 9). Cercus one-segmented (Figs 13,
 17) ***Greenomyia baikalica* Zaitzev, 1994**

The species

***Greenomyia baikalica* Zaitzev, 1994**

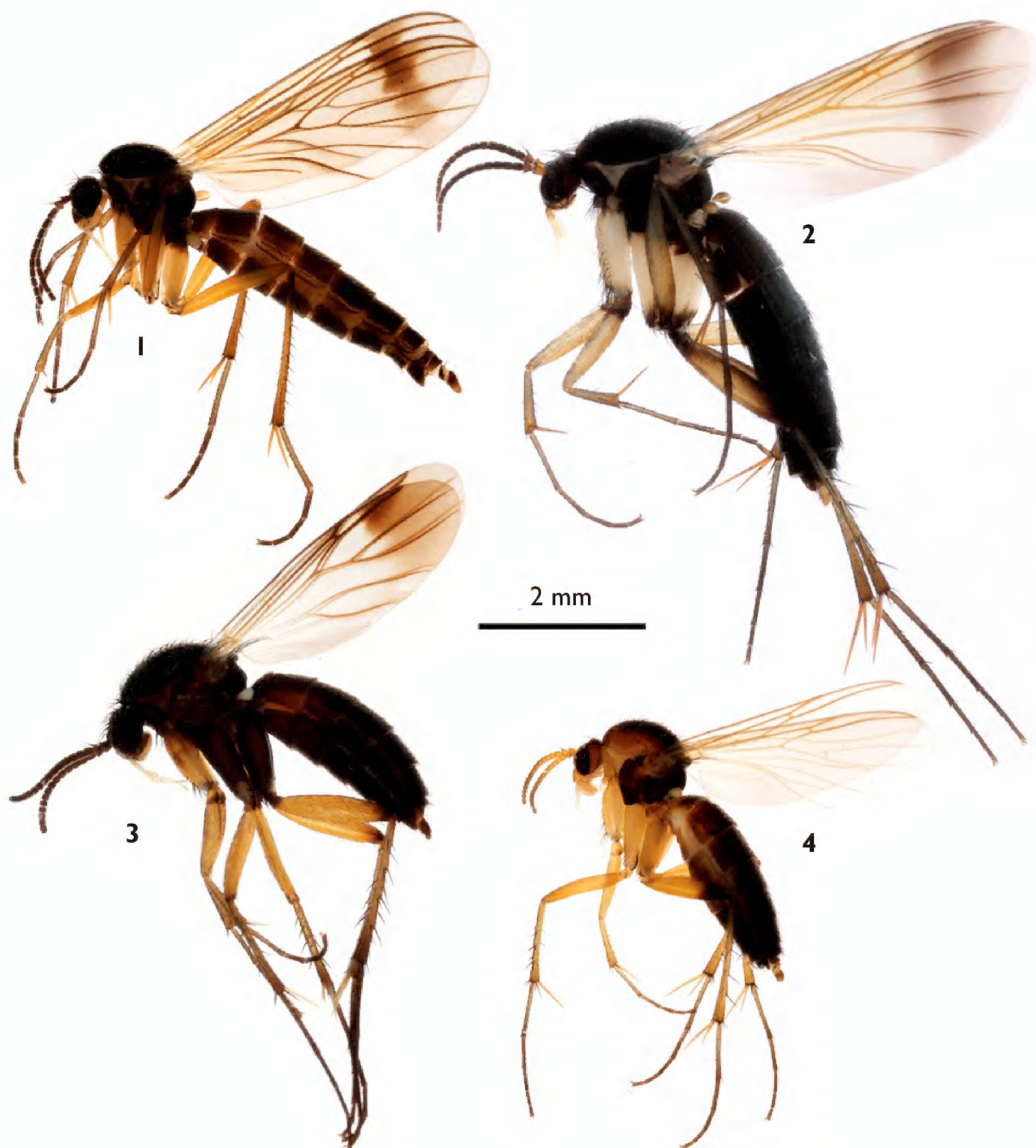
Figures 1, 5, 9, 13, 17, 21, 25

Material studied: SWEDEN. 1♀, Lu. Jokkmokk, Vuollerim, Bomyrberget, in forest 135 m.a.s.l., Malaise trap, 16.–18.VIII.2006 (K. Hedmark leg.) [IZBE, mounted from alcohol]; 1♀, Lu. Jokkmokk, Vuollerim, Bomyrberget, 135 m.a.s.l., Malaise trap, 18.–24.VIII.2007 (K. Hedmark leg.) [MZLU, in alcohol]. **FINLAND.** 1♂, Kn. Sotkamo, Urpovaara, window trap N2, 26.VIII.–11.IX.1997 (M. Kuussaari leg.) [IZBE, on pin].

Diagnostic characters. Female. Thorax blackish, abdomen brown. Legs yellow, cx_3 with small dark markings basally. All trochanters yellow, with small black apicoventral spots. Tibiae with dense brown setae. Scape and pedicel dark yellow, first flagellomere basally yellowish, rest of flagellum brown. Mouthparts pale yellow. Apical palpal segment 1.4–1.6 ($n=2$) times as long as penultimate segment. Wing with narrow preapical brownish band, gradually tapering towards hind margin. C terminating almost at apex of wing, R_5 distinctly arched (Fig. 9). Medial and cubital veins both reach wing margin, CuA_1 basally obsolete, brownish shade along posterior margin of CuA_2 . Terminalia brown, cercus one-segmented, yellow apically. Tergite IX larger than tergite VIII. Gonapophysis IX visible in lateral view, with wide pear-shaped medial incision apically. Sternite VIII deeply incised apicomediaally and moderately emarginated basally.

Male. Coloration and other non-terminal characters including palpi similar to female. Apical palpal segment is 1.4 ($n=1$) times as long as penultimate segment.

Remarks. Zaitzev (1994) described this species from Siberian material (Buryat Republic). Subsequently only a few specimens have been recorded from Norway, Sweden, Finland and Russian Karelia (cf. Kjærandsen et al. 2007; Fig. 29). Zaitzev (1994) figured the male terminalia whereas the female terminalia have not been figured earlier. Using the key by Zaitzev (1994) the studied females run to *G. baikalica* and they are also morphologically conspecific with material of both sexes collected simultaneously



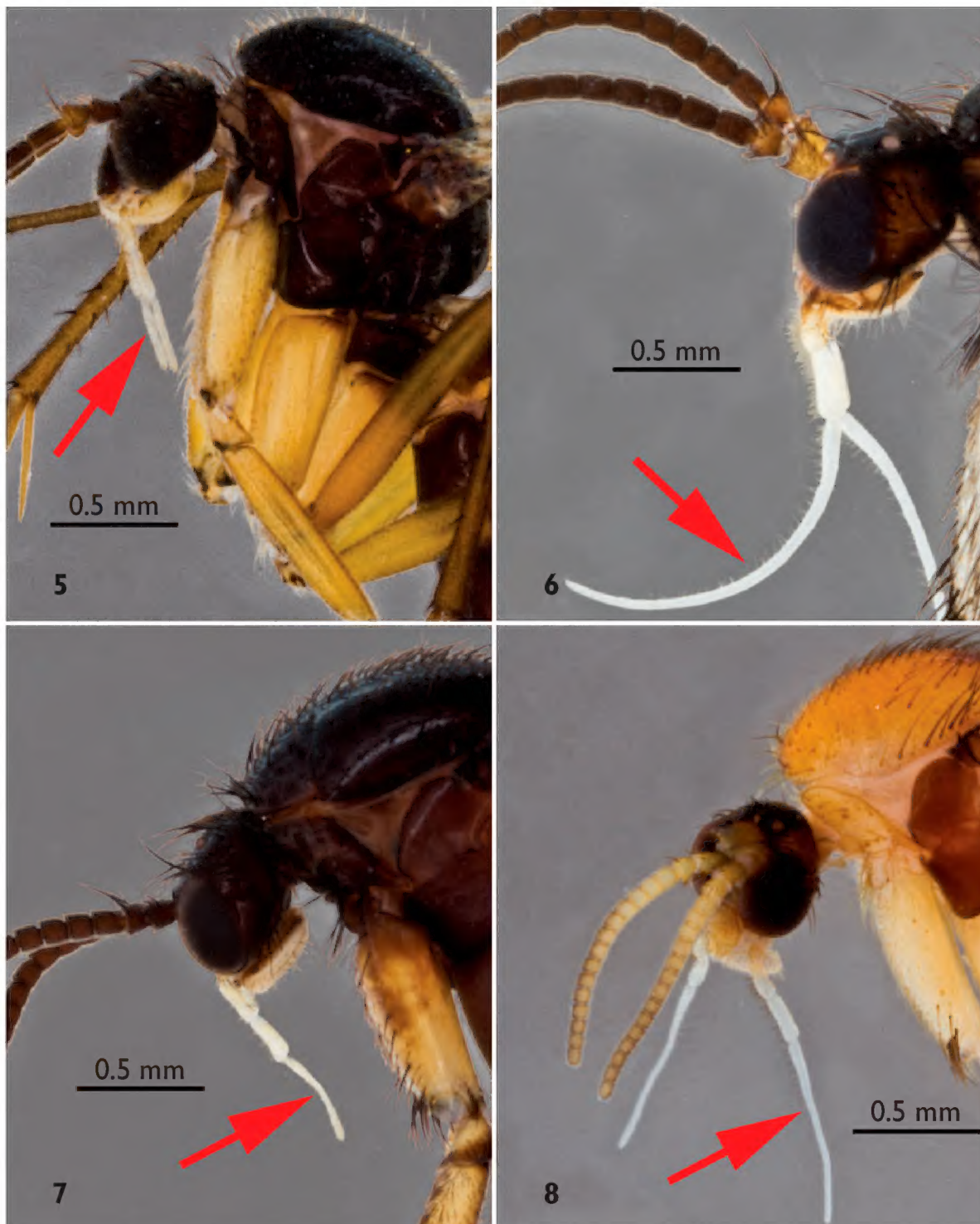
Figures 1–4. Habitus photos of European *Greenomyia* females. **1** *G. baikalica* **2** *G. borealis* **3** *G. mongolica* **4** *G. stackelbergi*.

in Russian Karelia (A. Polevoi, pers. comm.). The studied female specimens were collected in a boggy forest stand within a small (9 ha) protected remnant of semi-natural, mixed forest.

***Greenomyia borealis* (Winnertz, 1863)**

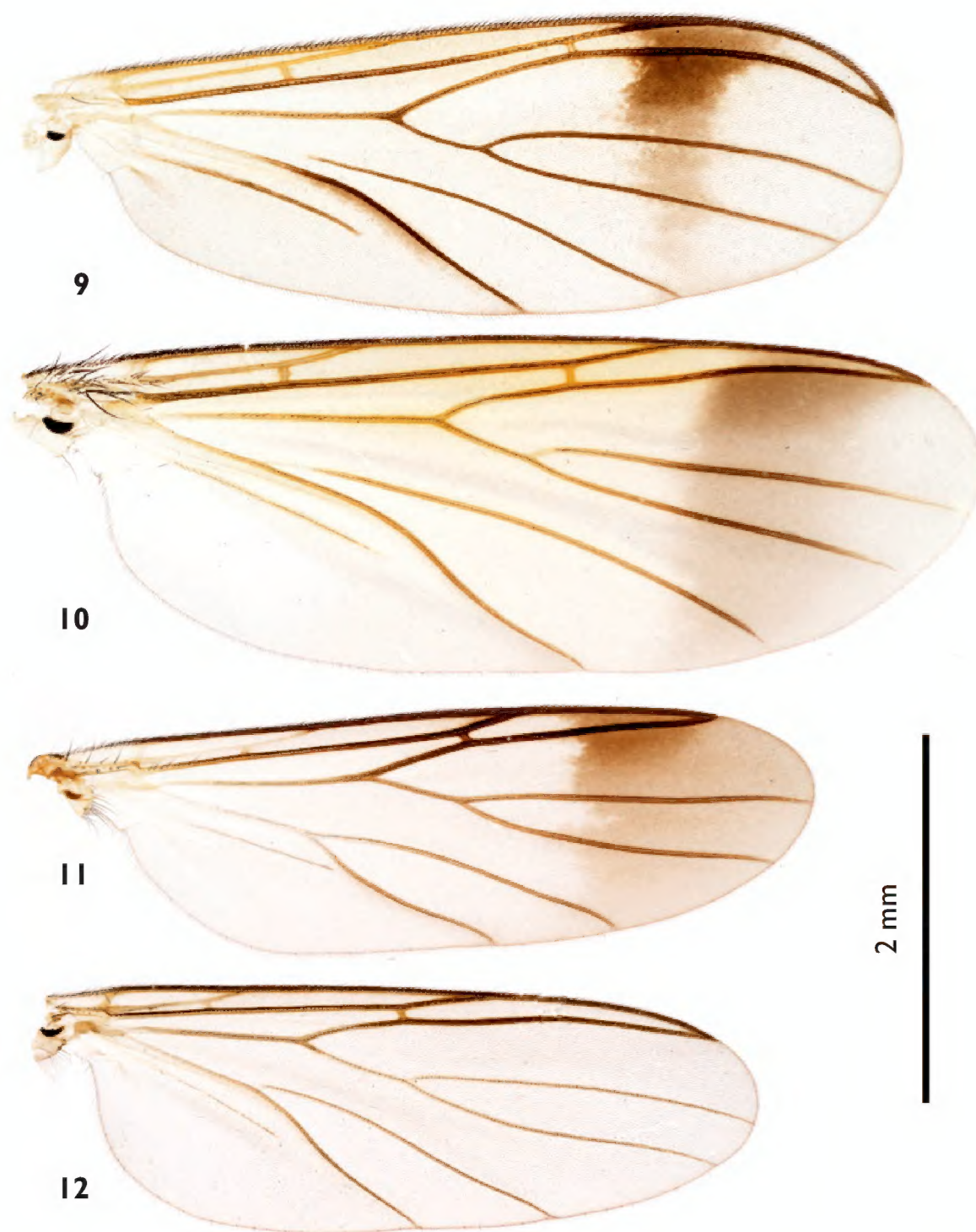
Figures 2, 6, 10, 14, 18, 22, 26

Material studied: SWEDEN. 2♂♂, SK, Lund and Lund, Abusa, undated (J. W. Zetterstedt leg.) [MZLU, on pins]; 1♀, ÖG, Valdemarsvik, Snäckevarp (Snäckehvarps



Figures 5–8. Head and palpi of European *Greenomyia* females, lateral view. **5** *G. baikalica* **6** *G. borealis* **7** *G. mongolica* **8** *G. stackelbergi*. Last palpal segment is indicated by a red arrow.

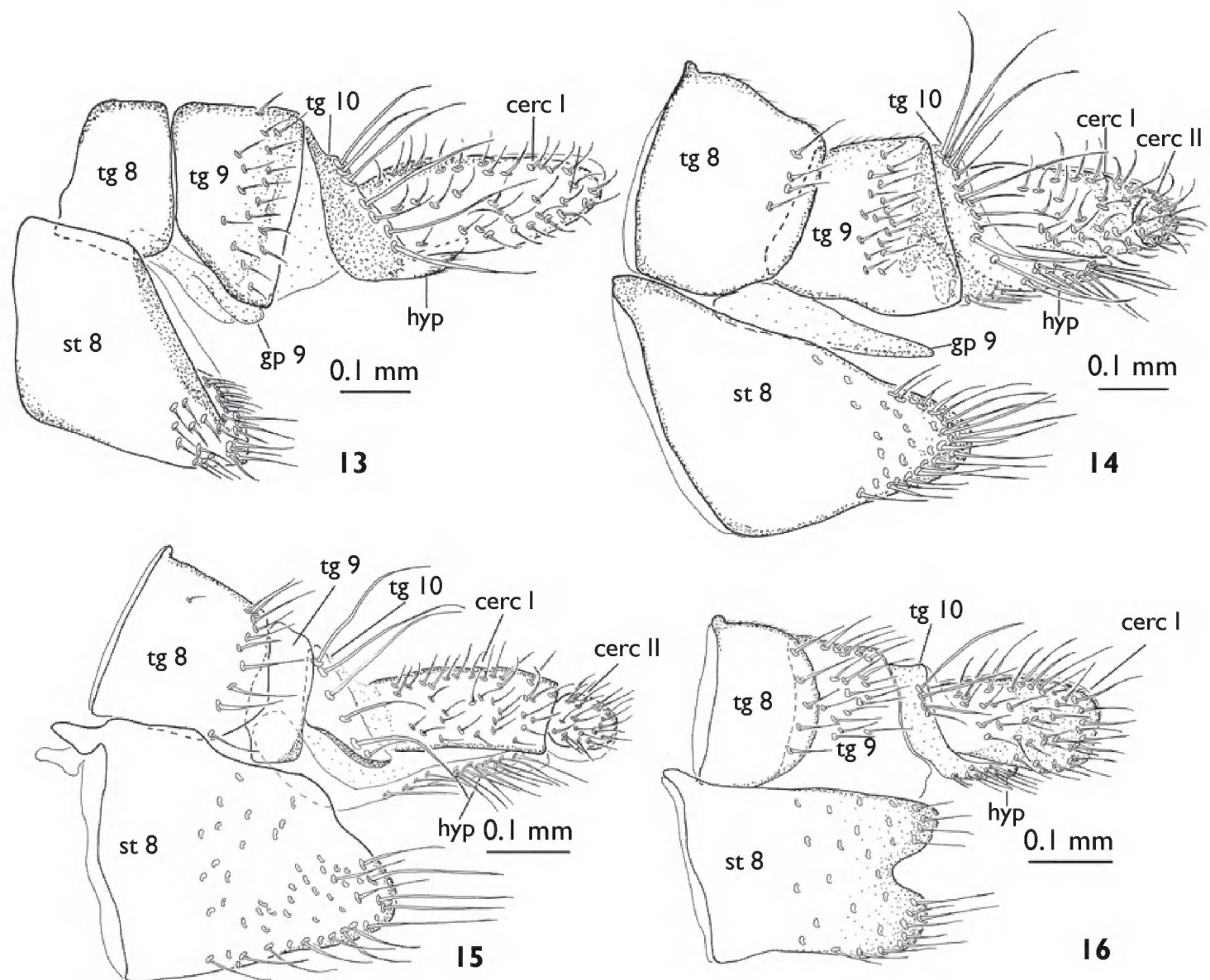
gästgifvaregård 1 km NE Gryt), 11 Aug 1825 (C. Stenhammar leg.) [MZLU, on pin]; 1♀, Lu. Jokkmokk, Vuollerim, in a garden, 105 m.a.s.l., Malaise trap, 8.–15.VIII.2008 (K. Hedmark and M. Karström leg.) [IZBE, mounted from alcohol]; **ESTONIA.** 1♂, Nigula NR, Haavapeaksi, sweeping, 12.VII.1998 (O. Kurina leg.) [IZBE, on pin]; 1♀, Tartu Marja 14, on window 21.VIII.2008 (O. Kurina leg.) [IZBE, on pin]. **GREECE.** 1♂ 2♀♀, Central Macedonia, Kerkini lakes area, village Vironia, Ramna site, 41°17'42.5"N, 023°11'33.1"E, 750 m.a.s.l., Malaise trap, 7.–13.VII.2008 (G. Ramel leg.) [IZBE, mounted from alcohol]; 1♂, Central Macedonia, Kerkini lakes area, village Vironia, Beabies site, 41°19'15.4"N, 023°13'39.6"E, 1150 m.a.s.l., Malaise trap, 21.–27.VII.2008 (G. Ramel leg.) [IZBE, mounted from alcohol]; 1♂ Central Macedonia, Kerkini lakes area, village Neo Petritsi, Midway site, 41°18'49.8"N,



Figures 9–12. Wings of European *Greenomyia* females. **9** *G. baikalica* **10** *G. borealis* **11** *G. mongolica* **12** *G. stackelbergi*.

023°16'35.6"E, 750 m.a.s.l., Malaise trap, 23.–29.VI.2008 (G. Ramel leg.) [IZBE, mounted from alcohol]. **KAZAKHSTAN.** 1♀, Alma-Ata, 13.–16.VI. 1824 (Kuzin leg.) [ZIN, on pin].

Diagnostic characters. Female. Thorax brown to blackish. Abdomen entirely brown or first two segments slightly lighter. Legs pale to yellow, except cx_2 and cx_3 with dark markings basally and apically, all trochanters brown and f_3 brown, with lateral parts lighter to yellow. Tibiae with dense brown setae. Scape and pedicel dark yellow, flagellomeres brown. Mouthparts yellow. Apical palpal segment 5.0–5.4 ($n=4$) times as long as penultimate segment. Wing with broad preapical brownish band, reaching hind margin but gradually paler. C terminating distinctly before apex of wing, R_5 straight to slightly sinuate (Fig 10). M_2 and CuA_1



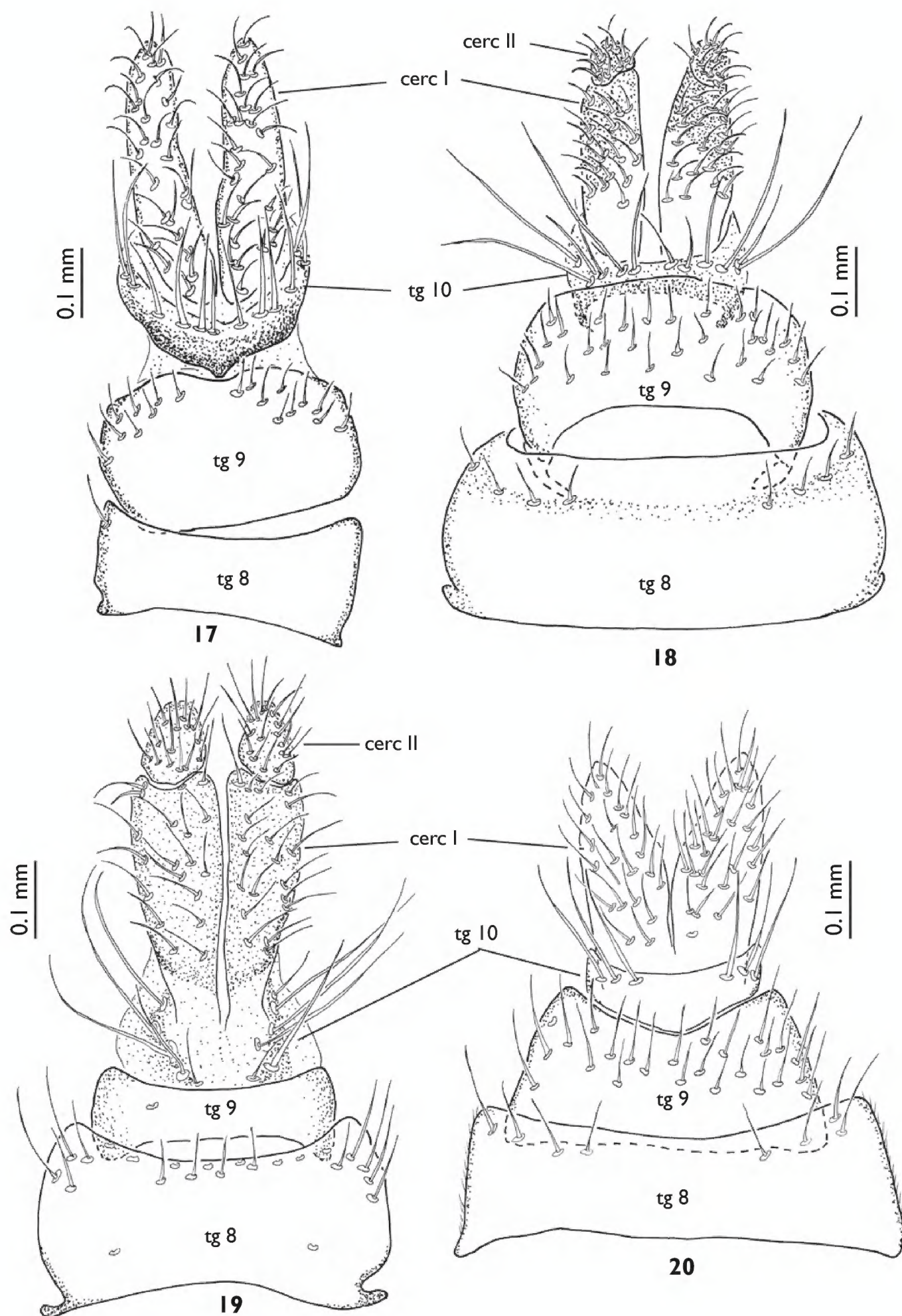
Figures 13–16. Female terminalia of European *Greenomyia* species, lateral view. **13** *G. baikalica* **14** *G. borealis* **15** *G. mongolica* **16** *G. stackelbergi*.

cerc = cercus; gp = gonapophysis; hyp = hypoproct; st = sternite; tg = tergite.

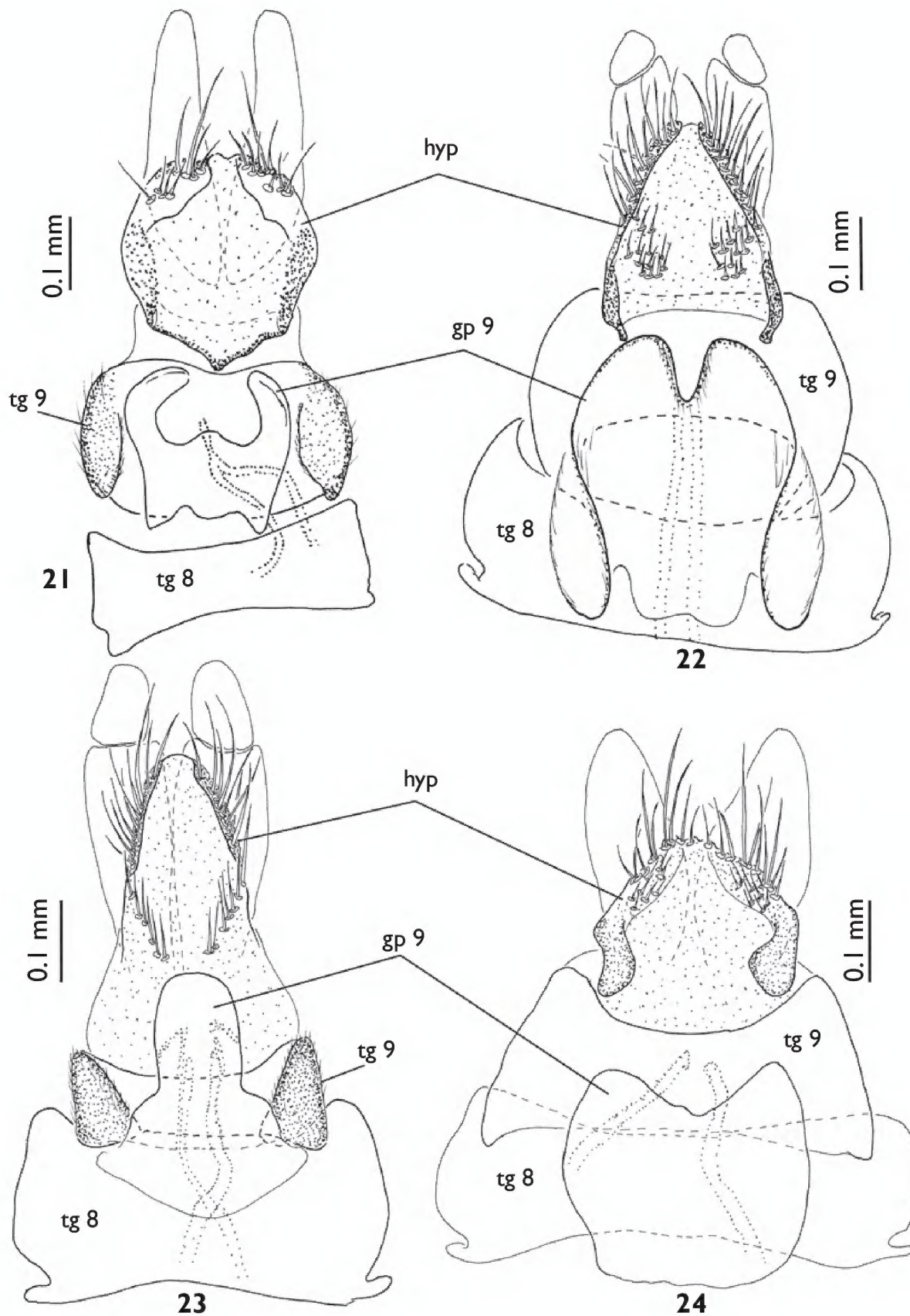
not reaching wing margin, CuA_1 basally obsolete or very weak. Terminalia brown; cercus yellow, two-segmented, apical segment small and partly fused with basal segment. Gonapophysis IX well sclerotized apically, visible in lateral view and with well developed narrow apical incision. Tergite VIII equal in size but slightly wider than tergite IX. Sternite VIII with medial incision apically and well emarginated basally.

Male. Coloration and other non-terminal characters including palpi similar to female. Apical palpal segment is 4.7–5.7 ($n=4$) times as long as penultimate segment.

Remarks. While studying the Swedish specimen from Vuollerim, it ran by the first attempt using the key by Zaitzev (1994) to *Neoclastobasis* because of the extra long last palpal segment and M_2 and CuA_1 not reaching the wing margin. The colouration of the studied specimen is, however, different and female terminalia lack strong spines on sternite VIII, being typical to all of the described *Neoclastobasis* species (Zaitzev 1982; JK and OK *pers. obs.* of *N. kamijoi*: 5♂♂ 2♀♀, South Korea, Sanan, I-li-Keumsan, [MNHN], 1♂ 3♀♀, Japan, Hokkaido, Sapporo [EIHU, MZLU]; *N. draskovicae*: paratypes, 1♂ 1♀ in MNHN, see Matile 1978). The dis-

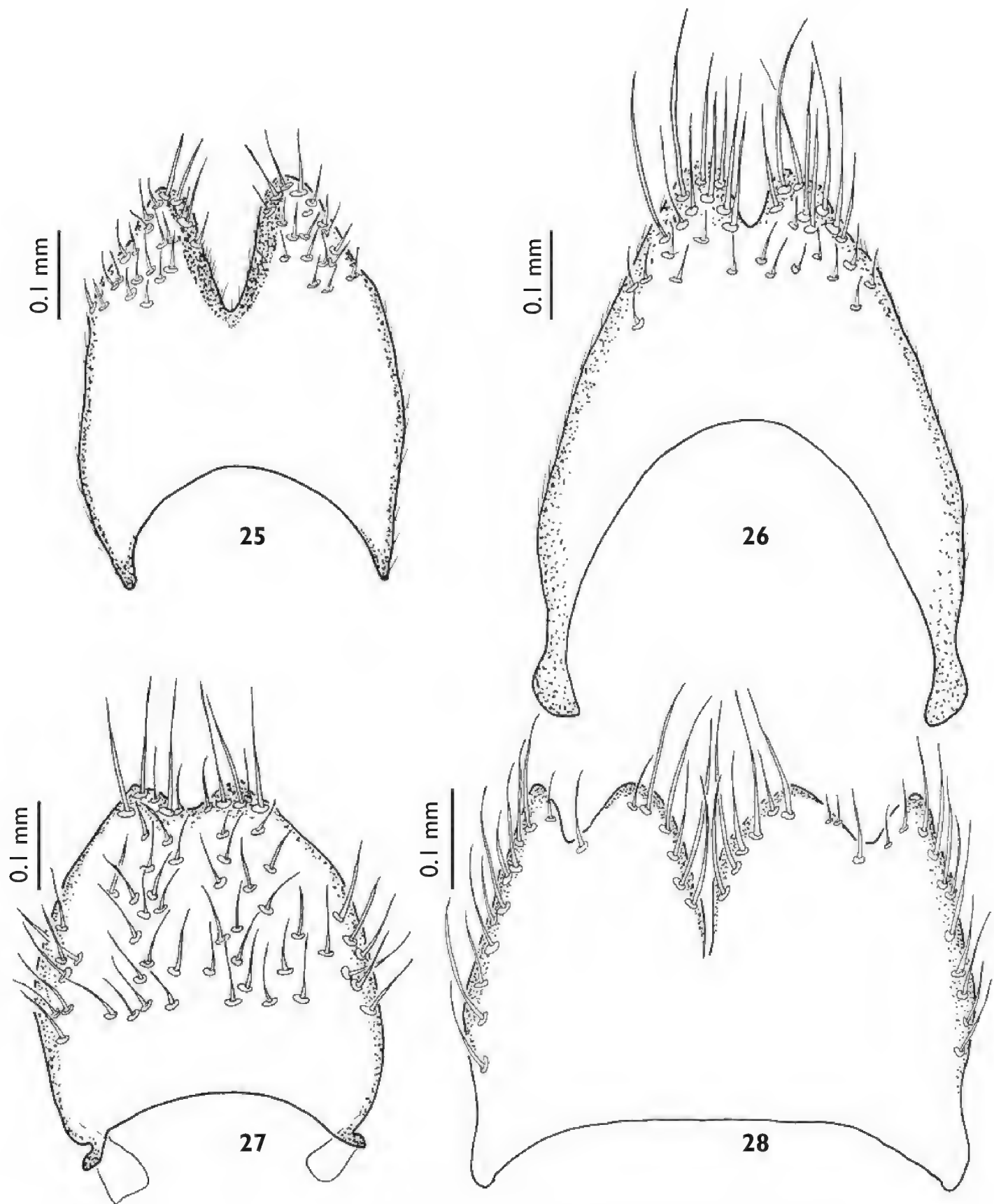


Figures 17–20. Female terminalia of European *Greenomyia* species, dorsal view. **17** *G. baikalica* **18** *G. borealis* **19** *G. mongolica* **20** *G. stackelbergi*.



Figures 21–24. Female terminalia of European *Greenomyia* species, ventral view, sternite VIII detached. **21** *G. baikalica* **22** *G. borealis* **23** *G. mongolica* **24** *G. stackelbergi*.

covery of a *Greenomyia* female, with similar size and coloration as the male of *G. borealis* in the same Malaise trap sample from northern Greece (Kerkini Lake area) allowed a safe association of the sexes. The females from Sweden, Estonia and Kazakhstan were further found to be conspecific with the Greek material of both sexes. According to Chandler et al. (2006), a male specimen from Greek mainland (Vikos Aios National Park) has mainly yellow coxae, while other European specimens of



Figures 25–28. Female terminalia of European *Greenomyia* species, ventral view of sternite VIII. **25** *G. baikalica* **26** *G. borealis* **27** *G. mongolica* **28** *G. stackelbergi*.

G. borealis have mainly dark coxae. This may represent an intraspecific variation, however, all specimens studied during the current investigation have coxae whitish yellow. The above-mentioned Estonian specimen represents the first record of *G. borealis* from the country. The female specimen from Vuollerim was collected in the same garden as *G. stackelbergi*. *G. borealis* was previously known only with two 19th century findings from southern Sweden.

***Greenomyia mongolica* Laštovka et Matile, 1974**

Figures 3, 7, 11, 15, 19, 23, 27

= *Greenomyia theresae* Matile, 2002, syn. n.*G. theresae* Kurina 2008a: 255, 270.

Type material studied: Paratype ♂ of *G. mongolica*: **MONGOLIA**. Central aimak, Tosgoni ovoo, 5–10 km N von Ulan-Baator, 1500–1700 m a.s.l., Exp. Dr. Z. Kaszab 1967 nr. 926, 19–24 Jul 1967 (Z. Kaszab leg.) [MNHN, JKJ-SPM-011843, on pin]

Holotype ♂ of *G. theresae*: **ITALY**. Aosta, Champlong, Dessus, 1000 m a.s.l., “courant sur Feuille de Frêne - 2m -”, 26 Aug 1997 (L. Matile leg.) [MNHN, JKJ-SPM-011844, on pin].

Other material studied: **SWEDEN**. 1♂, SÖ, Stockholm, Skarpnäck, Skarpa by, 13.VII.–4.X.2003 (B. Viklund leg.) [MZLU, in alcohol]. **GERMANY**. 1♂, D. Oberpfals, NM Main-Donau-Kanal (Proj. Warncke), 12.IX.–5.X.1988 (S. Blank leg.) [No. 30132 in ZSM, in alcohol]. **RUSSIA**. 2♂♂ 3♀♀, Nikolsk-Ussur, 29.VII.1926 (Kuznetzov leg.) [ZIN, on pins]. **ESTONIA**. 1♀, Kääriku, 5.X.1985 (H. Remm leg.) [IZBE, on pin]; 1♀, Luunja, 20.X.1996, on the house wall (O. Kurina leg.) [IZBE, on pin]; 6♂♂ 3♀♀, Karilatsi near Tartu, bait traps, 19.–28.VIII.2005 and 04.–25. IX.2005 (T. Tammaru leg.) [IZBE, on pins]. **HUNGARY**. 38♂♂ 3♀♀, 10 km S Eger, 47°49'11"N, 020°21'37"E, 20 Aug 1989 (R. Danielsson leg.) [MZLU, on pins]. **ITALY**. 1♂, Aosta valley, Verrayes, Promellian, 1200 m.a.s.l., sweeping, 17.VI.2007 (V. Soon leg.) [IZBE, on pin]; 1♀, Siena, 6.V.2007 (A. Selin leg.) [Coll. Selin, on pin]; 3♂♂, Trentino-Alto Adige, Prov. Bolzano, Parco Nazionale dello Stelvia, Sulden Valley near Schmelz southwest of Prad, 46°36'42.1"N, 010°34'35.6"E, 940 m.a.s.l., 5.IX.–14.X.2005 (J. Ziegler and C. Lange leg.) [1♂ in ZMHB, 2♂♂ in IZBE, mounted from alcohol]. **GREECE**. 1♂ 2♀♀, Central Macedonia, Kerkini lakes area, village Vironia, Beabies site, 41°19'15.4"N, 023°13'39.6"E, 1150 m.a.s.l., Malaise trap, 30.VI.–6.VII.2008 (G. Ramel leg.) [IZBE, mounted from alcohol]; 6♀♀, Central Macedonia, Kerkini lakes area, village Vironia, Ramna site, 41°17'42.5"N, 023°11'33.1"E, 750 m.a.s.l., Malaise trap, 23.–29.VI.2008 (G. Ramel leg.) [IZBE, mounted from alcohol].

Diagnostic characters. Female. Thorax dark brown to blackish. Abdomen entirely blackish brown or first three segments slightly paler laterally. cx_1 entirely yellow or darkened in basal half, cx_2 and cx_3 entirely dark brown to black. Fore trochanter yellow basally, brown apically. Mid and hind trochanters brown. f_1 and f_2 yellow, f_3 yellow with brown apical fifth. Tibiae yellow, apically slightly darkened, with dense brown setae. Scape, pedicel and flagellomeres brown. Mouthparts pale yellow. Apical palpal segment 1.8–2.2 (n=5) times as long as penultimate segment. Wing tip shaded on about apical third, with darkened area along fore margin. All veins reach wing margin, M_2 sometimes basally obsolete or very weak, A_1 ending close to, sometimes fused into base of CuA_2 . Terminalia brown. Cercus distinctly two-segmented, apical segment small, ovate. Gonapophysis IX membranous, widely protruding apically, not visible

in lateral view. Tergite VIII larger than tergite IX. Sternite VIII apically with shallow medial incision, moderately emarginated basally.

Male. Coloration and other non-terminal characters including palpi similar to female. Apical palpal segment is 1.7–2.1 (n=5) times as long as penultimate segment.

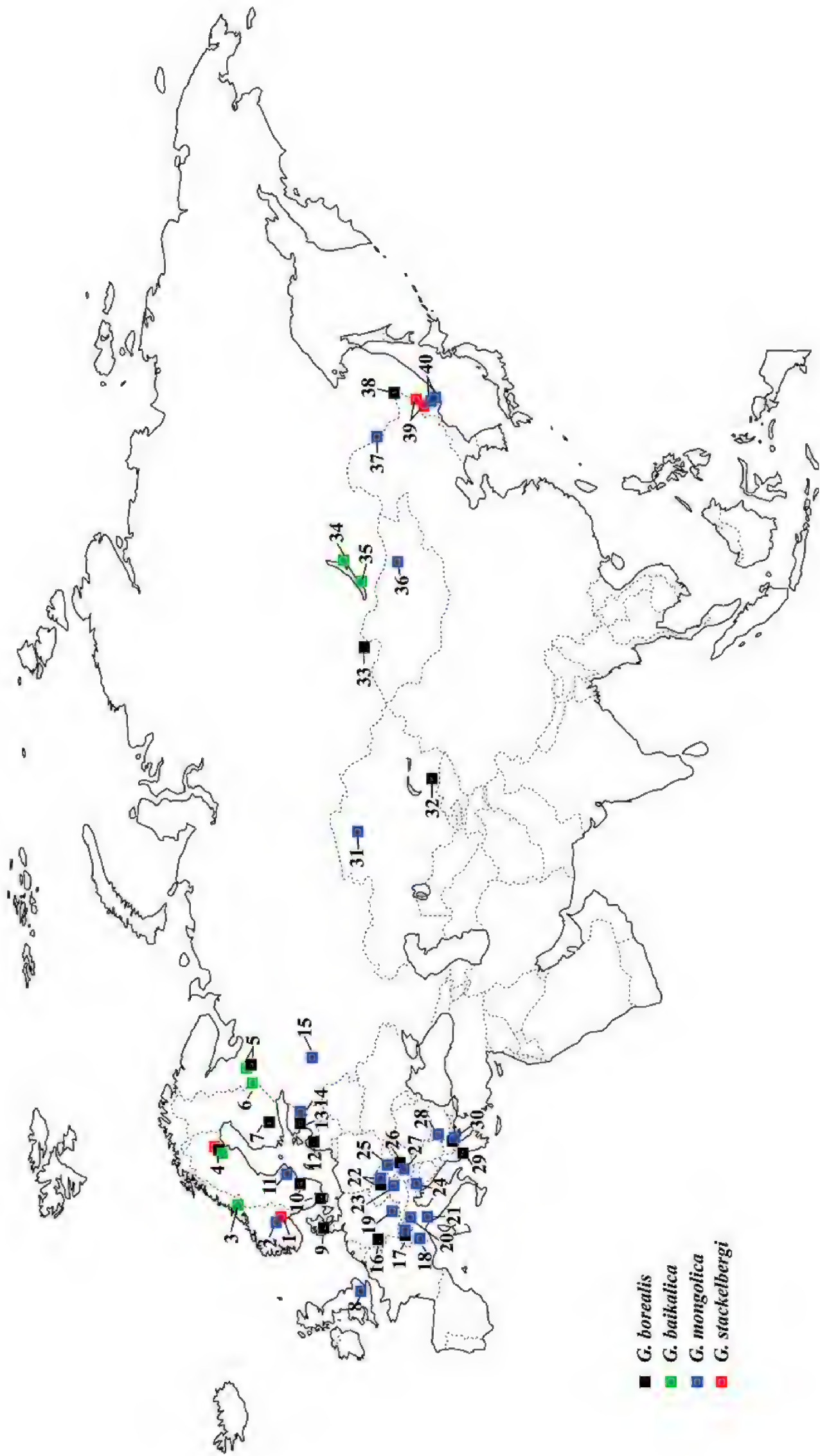
Remarks. This species was originally described by Laštovka and Matile (1974) based on Mongolian material and subsequently widely recorded in Europe. Chandler (2005) did not include *G. mongolica* in the European list and assigned all records to *G. theresae*, a species described from northern Italy by Matile (2002). Careful comparisons of type material of both species at MNHN in Paris (independently undertaken by two of the authors, OK and JK; the holotype of *G. mongolica* deposited in the Hungarian Natural History Museum was not available for the study) did not indicate any substantial differences in their male terminalia. The minor diagnostic characters as indicated in the original description and illustrations by Matile (2002) are liable to different angles of views only. Consequently we have come to the conclusion that *G. theresae* at present state of knowledge must be treated as a junior synonym of *G. mongolica* and that all published records in Europe should rather be associated with the latter. In addition to the studied type material, we also compared the terminalia of female specimens from the Russian Far East, Estonia and northern Greece without finding any reliable differences. Moreover, Papp (2000) confirmed conspecificity when he compared central European material from Hungary with the Mongolian type material. Male terminalia are figured by Laštovka and Matile (1974) and subsequently by Matile (2002), while female terminalia have previously been figured by Zaitzev (1982) and Kurina (1997). Our association of males and females are based on multiple simultaneous findings in trap samples (see above) that agrees with previous descriptions of the female. In the Pre-Balkan mountain range in Bulgaria, the species has been collected in xerothermic oak forest (Bechev 2000). The species was quite common in samples taken in a bait trap, operated on the basis of a mixture of fermenting sugar and red wine, in southern Estonia (see also Kurina 2006). The above-mentioned specimens from Greece are the first records from the country.

***Greenomyia stackelbergi* Zaitzev, 1982**

Figures 4, 8, 12, 16, 20, 24, 28

Type material studied: Holotype ♂, **RUSSIA**. Primorskiy Terr., Santaheza, 07.VII.1927 (A. Stackelberg leg.) [ZIN, on pin].

Other material studied: **SWEDEN**. 4 ♀♀, Lu. Jokkmokk, Vuollerim, in garden, 105 m.a.s.l., Malaise trap, 11.VIII.–19.IX.2003 (K. Hedmark and M. Karström leg.); 1♂, the same locality, Malaise trap 11.–19.VIII.2004; 25♂♂ 15♀♀, the same locality, Malaise trap 11.VIII.–7.X.2005; 11♂♂ 4♀♀, the same locality, Malaise trap 11.VIII.–22.IX.2006; 18♂♂ 14♀♀, the same locality, yellow pan-trap VII–08.X.2006; 17♂♂ 7♀♀, the same locality, yellow pan-trap 16.VI.–20.VII.2007; 14♂♂ 4♀♀, the same



locality, Malaise trap 12.VIII.–28.IX.2007; 9♂♂ 3♀♀, the same locality, window trap VI–11.IX.2007; 4♂♂ 1♀, the same locality, Malaise trap 13.–27.VI.2008; 1♀, the same locality, yellow pan-trap 19.VI.2008; 1♂, the same locality, window trap 1.VI.–1.VII.2008. In total 153 specimens: 100♂♂ 53♀♀, [most in Coll. Hedmark, some in IZBE and MZLU, most of the material preserved in alcohol, while some specimens are mounted from alcohol to pins or slide mounted].

Diagnostic characters. Female. Thorax bi-coloured; mesonotum yellow with variably developed black thoracic stripes; pronotum and propleuron yellow, other pleural parts brown to blackish. Abdominal sternites I–IV entirely yellow or slightly brownish; tergites of first four segments bi-coloured: basally yellow, apically brown (in a few occasions first four tergites entirely brown). Legs all yellow except dark brown band on apical fourth of hind femur. Tibiae densely covered with brown setulae. Scape, pedicel, and 3–5 flagellomeres yellow, rest of flagellum light brown. Mouthparts yellow. Apical palpal segment 4.1–4.4 (n=5) times as long as penultimate segment. Wing hyaline with slight yellowish tinge, all veins reach wing margin, M_1 and CuA_2 basally obsolete or very weak. Terminalia brown, cercus one-segmented, apically yellow. Gonapophysis IX membranous, subsquare with shallow incision apically, not visible in lateral view. Tergite VIII wider than tergite IX. Sternite VIII medially with deep and narrow incision, lateral incisions more shallow.

Male. Coloration and other non-terminal characters similar to female. The apical palpal segment is 4–5 (n=5) times as long as penultimate segment.

Remarks. Besides its peculiar distribution (see Fig. 29), *G. stackelbergi* is unique among the four studied species in having vivid yellowish colouration and hyaline wings. It was described from South Primorje in the Russian Far East (Zaitzev 1982) and has subsequently been recorded only from two semi-urban localities in the Nordic region: the single locality in Swedish Lapland (present material, Kjærandsen et al. 2007) and from one locality in the capital of Norway, Oslo (Søli and Kjærandsen 2008). Eight years of collecting (2002–2009) with Malaise traps, yellow pan-traps and window traps near a compost in the garden of one of the authors (MK) yielded 153

Figure 29. Known records of European *Greenomyia* species.

G. borealis (black squares): 4 original data 5 Polevoi 2000 (marked with a question mark) 7 Chandler 2005 9 Chandler 2005 10 Kjærandsen et al. 2007, original data 12 Lackschewitz 1937 13 original data from two localities 16 Winnertz 1863 17 Chandler 1998, 2005 (doubtful) 22 Ševčík and Košel 2009 26 Ševčík and Papp 2001 29 Chandler et al. 2006 30 original data 32 Zaitzev 1982, 1994 33 Zaitzev 1982, 1994 38 Zaitzev 1982, 1994.

G. baikalica (green squares): 3 Gammelmo and Søli 2006 4 Kjærandsen et al. 2007 5 Polevoi 2000 (three different localities) 6 Polevoi 2001 34 Zaitzev 1994 35 Zaitzev 1994.

G. mongolica (blue squares): 2 Søli et al. 2009 8 Chandler 2010 (seven different localities) 11 Kjærandsen et al. 2007 14 Kurina 1997 15 Zaitzev 1994 17 Matile 2002 (as *G. theresae*) 18 Matile 2002 (as *G. theresae*) 19 Caspers 1996 20 Kurina 2008a (as *G. theresae*) 21 original data 22 Martinovský and Ševčík 1998, Martinovský and Barták 2000 23 Plassmann 1996 24 Matile 2002 (as *G. theresae*) 25 Ševčík and Martinovský 1999 27 Papp 2000 (several different localities), original data 28 Bechev 1989 30 original data 36 Laštovka and Matile 1974 37 Zaitzev 1982, 1994 40 Zaitzev 1982, 1994.

G. stackelbergi (red squares): 1 Søli and Kjærandsen 2008 4 Kjærandsen et al. 2007 39 Zaitzev 1982, 1994.

specimens, indicating rise and decline of a small population. None was collected in the first and the last year, while four in 2003, one in 2004, 40 in 2005, 47 in 2006, 54 in 2007 and seven in 2008. The flight activity lasted almost the whole vegetation season, from the middle of June (in 2007) to the beginning of October (in 2004). A garden compost is the supposed microhabitat for this population of *G. stackelbergi* and its origin should be somewhere in the surroundings. A close potential natural habitat could be the Vuollerim ravine a few hundred meters away. Waste from picked forest fungi might be another possibility.

Discussion

Species descriptions of fungus gnats are largely based and depending on characters in the male terminalia. Females are often ignored in taxonomic reviews and only a few generic reviews cover all or the majority of associated females (e.g. Söli 1997; Martinsen and Söli 2000; Kjærandsen 2006, 2009). Still, females usually have distinctive yet less pronounced characters in their terminalia. In the case of the few European *Greenomyia* species we found it fairly easy to safely associate the females based on body characters such as colouration patterns, wing shape and venation details shared between the sexes, and the associations were further strengthened by co-occurrence in multiple trap samples.

Our study of *Greenomyia* revealed that the diagnostic characters used to distinguish *Greenomyia* and *Neoclastobasis* in keys (e.g. Söli et al. 2000) does not hold up to a closer scrutiny, especially when both sexes are considered. Both sexes of *G. borealis* have wings where M_2 and CuA_1 end slightly before the wing margin, and both sexes of *G. borealis* and *G. stackelbergi* have prolonged apical segment of their palps. These characters are akin to those used to diagnose *Neoclastobasis*. Yet, although the three known species of *Neoclastobasis* are very similar to *Greenomyia* in general appearance they show distinctive features in their terminalia that separate them from *Greenomyia*. In *Greenomyia* the dorsal branch of the male gonostylus always has two distinct combs of blunt spines on an otherwise bare inner surface. In *Neoclastobasis* the entire inner surface is covered with short blunt setae and a single row of larger spines is situated basally. Females of *Neoclastobasis* have a few short and strong spines along the apical margin of sternite VIII, which are never found in *Greenomyia*. We think *Greenomyia* and *Neoclastobasis* may prove to be monophyletic sistertaxa, but pending genetic studies and a better definition of the entire Leini clade we leave the question whether they deserve to retain their status as separate genera or could be joined into one. In the meantime separating the two genera must rest entirely on differences in their terminalia as described above.

Acknowledgements

OK was funded by grant 7558 of the Estonian Science Foundation, JK by the Swedish Taxonomy Initiative. We are grateful to the European Commission's Research Infrastructure for funding the study visits to MNHN via the SYNTHESYS programme (FR-TAF-956 for JK and FR-TAF-5005 for OK). We are much obliged to the curators M. Baylac (MNHN), C. Daugeron (MNHN), E. Nartshuk (ZIN), W. Schacht (ZSM) and J. Ziegler (ZMHB) for the opportunity to work with the collections and for loans of material. A. Polevoi (Petrozavodsk, Russia), in addition to valuable comments and information, is thanked for assistance in determining the first specimens of *G. stackelbergi* and for providing a male of *G. baikalica* for study. A. Selin (Tallinn, Estonia), J. Jakovlev (Vantaa, Finland) and G. Ramel (Kerkini, Greece) helped with material for study. Lars Hedström (Uppsala, Sweden) is acknowledged for help with interpreting an old specimen label of *G. borealis*. Peter Kerr (Sacramento, USA) and an anonymous reviewer are thanked for their comments and suggestions on the manuscript.

References

- Amorim DS, Rindal E (2007) Phylogeny of the Mycetophiliformia, with proposal of the subfamilies Heterotrichinae, Ohakuneinae, and Chiletrichinae for the Rangomaramidae (Diptera, Bibionomorpha). *Zootaxa* 1535: 1–92.
- Bechev D (1989) New species Mycetophiloidea (Diptera) for Bulgarian fauna. III. *Travaux Scientifiques d'Universite de Plovdiv, Biologie* 27 (6): 153–159. [in Bulgarian]
- Bechev D (2000) Seasonal activity and vertical distribution of fungus gnats (Diptera: Sciaroidea, excluding Sciaridae) in the western and the central parts of Stara Planina ridge (Bulgaria). *Travaux Scientifiques d'Universite de Plovdiv, Animalia* 36 (6): 51–66.
- Becker T (1908) Dipteren der Kanarischen Inseln. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 1908: 1–180.
- Brunetti E (1912) The Fauna of British India, including Ceylon and Burma. Diptera Nemato-sera (Excluding Chironomidae and Culicidae). London, 581 pp., pls. I–XII.
- Caspers N (1996) Fundort Schöngesing, Oberbayern Die Pilzmücken (Diptera Sciaroidea: Bolitophilidae, Diadocidiidae, Ditomyiidae, Keroplatidae, Mycetophilidae). *Entomofauna* 17 (25): 385–396.
- Chandler PJ (1998) 20. Mycetophilidae. In: Merz B, Bächli G, Haenni JP, Gonseth Y (Eds) *Fauna Helvetica* 1. Diptera-Checklist. Schweizerische Entomologische Gesellschaft, 113–125.
- Chandler PJ (2005) Fauna Europaea: Mycetophilidae. In: de Jong H (Ed) *Fauna Europaea: Diptera, Nematocera*. Fauna Europaea, version 1.2. Available from: <http://www.faunaeur.org> [accessed 21.I.2010]
- Chandler PJ (2010) Fungus Gnats Recording Scheme. Newsletter 4. *Dipterists Forum*, Spring 2010: 1–7.

- Chandler PJ, Bechev D, Caspers N (2006) The fungus gnats (Diptera: Bolitophilidae, Diadocidiidae, Ditomyiidae, Keroplatidae and Mycetophilidae) of Greece, its islands and Cyprus. *Studia Dipterologica* 12: 255–314.
- Chandler PJ, Ribeiro E (1995) The Sciaroidea (Diptera) (excluding Sciaridae) of the Atlantic Islands (Canary Islands, Madeira and the Azores). *Boletim do Museu Municipal do Funchal (Historia Natural), Suplemento No. 3*: 1–170.
- Edwards FW (1925) British Fungus-Gnats (Diptera, Mycetophilidae). With a revised Generic Classification of the Family. *Transactions of the Entomological Society of London* 73 (1924): 505–670.
- Evenhuis NL (2009) The insect and spider collections of the world website. Available at: <http://hbs.bishopmuseum.org/codens/> [accessed 13.II.2010]
- Gammelmo O (2004). Classification of Mycetophilidae (Diptera, Sciaroidea). *Norwegian Journal of Entomology* 51: 145–149.
- Gammelmo O, Søli GEE (2006) Norwegian fungus gnats of the family Mycetophilidae (Diptera, Nematocera). *Norwegian Journal of Entomology* 53: 57–69.
- Kjærandsen J (2006) Review of fungus gnats of the genus *Tarnania* Tuomikoski, with a phylogeny of the *Rymosia* s. l. genus group (Diptera: Mycetophilidae). *Insect Systematics and Evolution* 37: 121–148.
- Kjærandsen J (2009) The genus *Pseudexechia* Tuomikoski re-characterized, with a review of European species (Diptera: Mycetophilidae). *Zootaxa* 2056: 1–45.
- Kjærandsen J, Hedmark K, Kurina O, Polevoi A, Økland B, Götmark F (2007) Annotated checklist of fungus gnats from Sweden (Diptera: Bolitophilidae, Diadocidiidae, Ditomyiidae, Keroplatidae and Mycetophilidae). *Insect Systematics and Evolution Supplement No. 6*: 1–128.
- Kurina O (1997) *Greenomyia mongolica* Lastovka et Matile, 1974 (Diptera, Mycetophilidae) found in Estonia. *An international Journal of Dipterological Research* 8: 69–71.
- Kurina O (2006) A review of Estonian wood gnats (Diptera: Anisopodidae). *Sahlbergia* 11: 18–22.
- Kurina O (2008a) Sciaroidea excl. Sciaridae. In: Ziegler J (Ed) *Diptera Stelviana. A dipterological perspective on a changing alpine landscape. Volume 1. Studia Dipterologica. Supplements* 16: 245–293.
- Kurina O (2008b) *Cluzobra matilei* sp. n. from French Guyana, with notes on congeners (Diptera: Mycetophilidae). *Zootaxa* 1874: 63–68.
- Lackschewitz P (1937) Die Fungivoriden des Ostbaltischen Gebietes. *Arbeiten des Naturforscher Verein zu Riga (N.F.)* 21: 1–47
- Laštovka P, Matile L (1974) Mycetophilidae (Diptera) de Mongolie. *Acta Zoologica Academiae Scientiarum Hungaricae* 20(1/2): 93–135.
- Martinsen L, Søli GEE (2000) Description of females of three species of *Ectrepesthoneura* Enderlein (Diptera, Mycetophilidae). *Norwegian Journal of Entomology* 47: 137–147.
- Martinovský J, Ševčík J (1998) Bolitophilidae, Diadocidiidae, Ditomyiidae, Keroplatidae, Macroceridae, Mycetophilidae. *Folia Facultatis Scientiarum Naturalium Universitatis Masarykianae Brunensis Biologia*. 99: 41–47.
- Martinovský J, Barták M (2000) Mycetophilidae. In Barták M, Vaňhara J (Eds) *Diptera in an industrially affected region (North-Western Bohemia, Bílina and Duchcov environs)* I.

- Folia Facultatis Scientiarum Naturalium Universitatis Masarykianae Brunensis, Biologia 104: 65–71.
- Matile L (1978) Description d'un *Neoclastobasis* nouveau de Hongrie et remarques sur divers Leiini (Diptera: Mycetophilidae). Folia Entomologica Hungarica, S. N. 31: 167–172.
- Matile L (2002) Notes sur le genre *Greenomyia* Brunetti et description d'une espece nouvelle des Alpes et des Apennins (Diptera: Mycetophilidae). Annales de la Société Entomologique de France (Nouvelle série) 38 (1–2): 13–16.
- Meigen JW (1818) Systematische Beschreibung der bekannten europäischen zweiflügeligen Insekten. Vol 1, Aachen , xxxvi+333pp.
- Ostroverkhova GP (1970) New data on Siberian fungus Gnats (Diptera, Mycetophilidae). Entomologicheskoe Obozrenie 49: 271–274. [In Russian]
- Papp L (2000) Pediciidae, Bolitophilidae, Keroplatidae, Mycetophilidae and Dixidae: genera and species new to Hungary (Diptera). Folia Entomologica Hungarica 61: 219–231.
- Plassmann E (1996) Zur Kenntnis der Pilzmückenfauna Österreichs (Diptera: Nematocera: Mycetophilidae). Mitteilungen des internationalen Entomologischen Vereins 21 (3/4): 111–120.
- Polevoi AV (2000) Fungus gnats (Diptera: Bolitophilidae, Ditomyiidae, Keroplatidae, Diadocidiidae, Mycetophilidae) of Karelia. Petrozavodsk, 84pp. [In Russian]
- Polevoi A (2001) The study of forest Diptera fauna in Koitajoki area. In: Hokkanen TJ (Ed) Diversity studies in Koitajoki area (North Karelian Biosphere Reserve, Ilomantsi, Finland). Metsähallitus, Vantaa, 72–85.
- Rindal E, Søli GEE, Bachmann L (2009) On the systematics of the fungus gnat subfamily Mycetophilinae (Diptera): a combined morphological and molecular approach. Journal of Zoological Systematics and Evolutionary Research 47(3): 227–233.
- Sasakawa M (1964) Japanese Mycetophilidae V. Descriptions of three new species. Akitu 12(1): 1–4.
- Søli GEE (1997) On the morphology and phylogeny of Mycetophilidae, with a revision of *Coelosia* Winnertz (Diptera, Sciaroidea). Entomologica Scandinavica Supplement 50: 1–139.
- Søli G, Kjærandsen J (2008) Additions to the Norwegian fauna of fungus gnats (Diptera, Mycetophilidae). Norwegian Journal of Entomology 55: 31–41.
- Søli GEE, Vockeroth RJ, Matile L (2000) A.4. Families of Sciaroidea. In: Papp L, Darvas B (Eds) Contribution to a Manual of Palearctic Diptera. Appendix. Science Herald, Budapest, 49–92.
- Søli GEE, Rindal E, Hansen LO (2009) New records of fungus gnats for Norway (Diptera: Mycetophilidae). Norwegian Journal of Entomology 56: 69–73.
- Ševčík J, Košel V (2009) Mycetophilidae Newman, 1834. In: Jedlička L, Kúdela M, Stloukalová V (Eds) Checklist of Diptera of the Czech Republic and Slovakia. Electronic version 2. <http://zoology.fns.uniba.sk/diptera2009> [accessed 07.II.2010]
- Ševčík J, Martinovský J (1999) Faunistic records. Keroplatidae, Mycetophilidae. In: Jedlička L (Ed) Dipterologica bohemoslovaca. Vol. 9. Slovak Entomological Society, Bratislava, 189–193.
- Ševčík J, Papp L (2001) Bolitophilidae and Mycetophilidae (Diptera): genera and species new to Hungary. Folia Entomologica Hungarica 62: 217–229.

- Vockeroth JR (1966) A method of mounting insects from alcohol. The Canadian Entomologist 98:69–70.
- Winnertz J (1863) Beitrag zu einer Monographie der Pilzmücken. Verhandlungen der Zoologisch-Botanischen Gesellschaft in Wien. 13: 637–964.
- Zaitzev AI (1982) *Greenomyia* and *Neoclastobasis*. Fungus gnats (Diptera, Mycetophilidae) of the USSR. Vestnik Zoologii 1982: 25–32. [in Russian]
- Zaitzev AI (1994) Fungus gnats of the fauna of Russia and adjacent regions. Part 1. Moscow, 288pp. [in Russian, with English summary]